Nori® Canine GLUT1/SLC2A1 ELISA Kit DataSheet

Glucose transporter 1 (or GLUT1), also known as solute carrier family 2, facilitated glucose transporter member 1 (SLC2A1), is a uniporter protein that in Canine GLUT1/SLC2A1s is encoded by the SLC2A1 gene.[2] GLUT1 facilitates the transport of glucose across the plasma membranes of mammalian cells.[3] Energy-yielding metabolism in erythrocytes depends on a constant supply of glucose from the blood plasma, where the glucose concentration is maintained at about 5mM. Glucose enters the erythrocyte by facilitated diffusion via a specific glucose transporter, at a rate about 50,000 times greater than uncatalyzed transmembrane diffusion. GLUT1 is responsible for the low-level of basal glucose uptake required to sustain respiration in all cells. Expression levels of GLUT1 in cell membranes are increased by reduced glucose levels and decreased by increased glucose levels. GLUT1 is also a major receptor for uptake of Vitamin C as well as glucose, especially in non-vitamin C producing mammals as part of an adaptation to compensate by participating in a Vitamin C recycling process. In mammals that do produce Vitamin C, GLUT4 is often expressed instead of GLUT1.[5] It is widely distributed in fetal tissues. In the adult it is expressed at highest levels in erythrocytes and also in the endothelial cells of barrier tissues such as the blood–brain barrier. GLUT1 behaves as a Michaelis-Menten enzyme and contains 12 membrane-spanning alpha helices, each containing 20 amino acid residues. Mutations in the GLUT1 gene are responsible for GLUT1 deficiency or De Vivo disease, which is a rare autosomal dominant disorder.[6] This disease is characterized by a low cerebrospinal fluid glucose concentration (hypoglycorrhachia), a type of neuroglycopenia, which results from impaired glucose transport across the blood–brain barrier. GLUT1 is also a receptor used by the HTLV virus to gain entry into target cells.[7] Glut1 has also been demonstrated as a powerful histochemical marker for haemangioma of infancy.[8]

References

PRINCIPLE OF THE ASSAY

This is a quick ELISA assay that reduces time to 50% compared to the conventional method, and the entire assay only takes 3 hours. This assay employs the quantitative sandwich enzyme immunoassay technique and uses biotin-streptavidin chemistry to improve the performance of the assays. An antibody specific for Canine GLUT1/SLC2A1 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any GLUT1/SLC2A1 present is bound by the immobilized antibody. After washing away any unbound substances, a detection antibody specific for Canine GLUT1/SLC2A1 is added to the wells. Following wash to remove any unbound antibody reagent, a detection reagent is added. After intensive wash a substrate solution is added to the wells and color develops in proportion to the amount of GLUT1/SLC2A1 bound in the initial step. The color development is stopped and the intensity of the color is measured.

This package insert must be read in its entirety before using this product.

Storage Store at 4 °C. The kit can be used in 3 months.
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MATERIALS PROVIDED

<table>
<thead>
<tr>
<th>Description</th>
<th>Quantity</th>
<th>Description</th>
<th>Quantity</th>
<th>Description</th>
<th>Quantity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Antibody Precoated Plate</td>
<td>1</td>
<td>20 x PBS</td>
<td>1</td>
<td>Substrate Solution</td>
<td>1</td>
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<tr>
<td>Detection Antibody</td>
<td>1</td>
<td>20 x Assay buffer</td>
<td>1</td>
<td>Stop Solution</td>
<td>1</td>
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<tr>
<td>Conjugate</td>
<td>1</td>
<td>Reagent Diluent</td>
<td>1</td>
<td>DataSheet</td>
<td>1</td>
</tr>
<tr>
<td>Standard</td>
<td>3</td>
<td>MSDS/CoA</td>
<td>1</td>
<td>96-well plate sheet</td>
<td>1</td>
</tr>
</tbody>
</table>

Bring all reagents to room temperature before use.

**Reagent Preparations**

**Canine GLUT1/SLC2A1 Detection Antibody** (1 vial) – The lyophilized Detection Antibody should be stored at 4°C in a manual defrost freezer for up to 3 months, if not used immediately. Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. The vial contains sufficient Detection Antibody for a 96-well plate. Add 200 µL of sterile 1 x PBS to the antibody vial and vortex briefly and sit for 5 min. Take 200 µL of detection antibody to 10.5 mL of Reagent Diluent to make **working dilution of Detection Antibody** if the entire 96-well plate is used. If the partial antibody is used store the rest at -20°C until use.

**Canine GLUT1/SLC2A1 Standard** (3 vials) – The lyophilized Canine GLUT1/SLC2A1 Standard has a total of 3 vials. Each vial contains the standard sufficient for a 96-well plate. The non-reconstituted standard can be stored at 4°C for up to 3 months if not used immediately. Centrifuge for 1 min at 6000 x g to bring down the material prior to open the tube. Add 500 µL of 1 x Assay Buffer to one Standard vial to make the high standard concentration of 8 ng/ml. Vortex 20 sec and allow it to sit for 5 min prior to use. A seven-point standard curve is generated using 2-fold serial dilutions in the Assay Buffer, vortex 20 sec for each of dilution step.

**Conjugate** (53 µL) – Centrifuge for 1 min at 6000 x g to bring down the material prior to open the vial. The vial contains sufficient Conjugate for a 96-well plate. If the volume is less than 53 µL, add sterile 1 x PBS to reach 53 µL and vortex briefly. Make 1:200 dilution in Reagent Diluent. If the entire 96-well plate is used, add 53 µL of Conjugate to 10.5 mL of Reagent Diluent to make **working dilution of Conjugate** prior to the assay. The rest of undiluted Conjugate can be stored at 4°C for up to 3 months. DO NOT FREEZE.

**20 x PBS**, pH 7.3, 30 mL- Dilute to 1 x PBS with deionized distilled water and mix well prior to use.
**20 x Assay buffer**, 20 mL- Dilute to 1 x Assay buffer with 1 x PBS prior to use.
**Reagent Diluent**, 21 mL.
**Substrate Solution**, 10.5 mL.
**Stop Solution**, 5.5 mL.
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Assay Procedure

1. Lift the plate cover from the top left corner and cover the wells that are not used. Vortex briefly the samples prior to the assay. Add 100 µL of sample (such as plasma or serum) or standards per well and use duplicate wells for each sample. Cover the 96-well plate and incubate 1 hour at room temperature.

2. Aspirate each well and wash with 1 x Assay buffer, repeating the process two times for a total of three washes. Wash by filling each well with 1 x Assay buffer (300 µL) using a multi-channel pipette, manifold dispenser or auto-washer. Complete removal of liquid at each step is essential for good performance. After the last wash, remove any remaining Assay buffer by aspirating or by inverting the plate and blotting it against clean paper towels.

3. Add 100 µL of the working dilution of Detection Antibody to each well. Cover the plate and incubate 1 hour at room temperature.

4. Repeat the aspiration/wash as in step 2.

5. Add 100 µL of the working dilution of Conjugate to each well. Cover the plate and incubate for 20 minutes at room temperature. Avoid placing the plate in direct light.

6. Repeat the aspiration/wash as in step 2.

7. Add 100 µL of Substrate Solution to each well. Incubate for 5-20 minutes (depending on color development) at room temperature. Avoid placing the plate in direct light.

8. Add 50 µL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.

9. Determine the optical density of each well immediately, using a microplate reader set to 450 nm. If wavelength correction is available, set to 540 nm or 570 nm. If wavelength correction is not available, subtract readings at 540 nm or 570 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

Precaution and Technical Notes

1. It is critical to follow the procedure step by step otherwise appropriate color development may not occur as expected.

2. A standard curve should be generated for each set of samples assayed. Thorough mixing of the standard at each of the dilution steps is critical to ensure a normal calibration curve.

3. Plasma or serum sample should be diluted with equal volume of 1 x Assay Buffer and vortex for 1 min prior to assay. If the OD value still exceeds the upper limit of the standard curve, further dilution is recommended till it falls in the detection range and the dilution factor must be used for calculation of the concentration.

4. Conjugate contains enzyme, DO NOT mass up with Detection Antibody.

5. The Stop Solution is an acid solution, handle with caution.

6. This kit should not be used beyond the expiration date on the label.

7. A thorough and consistent wash technique is essential for proper assay performance.

8. Use a fresh reagent reservoir and pipette tips for each step.

9. It is recommended that all standards and samples be assayed in duplicate.

10. Avoid microbial contamination of reagents and buffers. This may interfere with the sensitivity of the assay.
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Calculation of Results
Average the duplicate readings for each standard, control, and sample and subtract the average zero (blank) standard optical density.

Create a standard curve by reducing the data using computer software capable of generating a four-parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the GLUT1/SLC2A1 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

The Standard Curve
The graph below represents typical data generated when using this Canine GLUT1/SLC2A1 ELISA Kit. The standard curve was calculated using a computer generated 4-PL curve-fit. For this case, a Bio-Rad iMark™ Microplate Reader and Microplate Manager 6 Software were used to generate this curve. The correlation coefficient ($r^2$) is 0.999-1.000.
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Specificity
The following recombinant Canine GLUT1/SLC2A1 proteins prepared at 1 ng/ml were tested and exhibited no cross-reactivity or interference. ApoA1, BMP1, BMP2, BMP3, BMP4, CCL4/MIP-1β, CRP, HSP27, IL-1β, IL-1ra, IL-2, IL-4, IL-5, IL-6, IL-8, IL-12, IL-15, IL-17C, IL-21, IL-23, IL-2R, IL6R, IFNγ, PDGF, PLA2G7, prolactin, TGFβ1, TGFβ2, TGFβ3, TLR1, TLR2, TLR3, TNF-α, TNF RI, TNF RII, VEGF.

Calibration
This kit is calibrated against a highly-purified yeast-expressed recombinant Canine GLUT1/SLC2A1.

Detection Range
125-8000 pg/ml

Assay Sensitivity
25 pg/ml

Assay Precision
Intra-Assay %CV: 5; Inter-Assay %CV: 8

For Research Use Only.

Related products
20 x PBS, Cat. GR103004-20
10 x ELISA Assay buffer, Cat. GR103028
10 x ELISA Reagent Diluent, Cat. GR103055
Universal Blocking Buffer, Cat. GR103005
2 x Recombinant Protein Stabilizer, Cat. GR03014-2
5 x Recombinant Protein Stabilizer, Cat. GR03014-5
ELISA G-Blue Substrate Solution, Cat. GR103021
Recombinant Canine GLUT1/SLC2A1
Canine GLUT1/SLC2A1 Detection Antibody
## Nori® Canine GLUT1/SLC2A1 ELISA Kit DataSheet

### Troubleshooting Guide

<table>
<thead>
<tr>
<th>Problem</th>
<th>Possible causes</th>
<th>Solution</th>
</tr>
</thead>
</table>
| Poor standard curve      | - Inaccurate pipetting  
- Improper standard curve                                                       | - Check pipettes  
- Check and use the correct dilution buffer  
- Vortex 30 sec for each of standard dilution steps                                      |
| Low signal               | - Improper preparation of standard, samples, detection antibody, and/or conjugate  
- Too brief incubation times  
- Inadequate reagent volume or improper dilution                                       | - Briefly spin down vials before opening. Reconstitute the powder thoroughly.  
- Ensure sufficient incubation time.  
- Check pipettes and ensure correct preparation.                                         |
| Large CV                 | - Inaccurate pipetting and mixing  
- Improper standard/sample dilutions.  
- Air bubbles in wells.                                                                  | - Check pipettes and ensure thorough mixing.  
- Use the correct dilution buffers  
- Remove bubbles in wells.                                                              |
| High background          | - Plate is insufficiently washed.  
- Contaminated assay buffer                                                            | - Review the datasheet for proper wash. If using a plate washer, ensure that all ports are unobstructed.  
- Make fresh assay buffer                                                               |
| No signal detected       | - The procedure was misconducted.                                                 | - Ensure the step-by-step protocol was correctly followed and no misstep was conducted.       |
| Low sensitivity          | - Improper storage of the ELISA kit  
- Stop solution                                                                         | - Store standards and detection antibody at -20°C after reconstitution, others at 4°C.  
Keep substrate protected from light.  
- Adding stop solution to each well before reading plate                                  |